

Plant Tissue Sampling

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Background

- Plant analyses are useful to diagnose nutritional problems and to monitor the fertilization program. Tissue testing is most effective when used together with soil testing ^[13].
- Nutrient concentrations change as plants grow and also differ between plant parts ^[6]. It is therefore important to sample specific plant parts at a particular growth stage (Table 1). For information on optimal nutrient concentrations at different growth stages, see the fertilization guidelines for the different crops (<http://apps.cdfa.ca.gov/frep/docs/Intro.html>)
- Archiving the results from the analyses allows tracking changes in the same field over time ^[6]. Plant analyses together with soil analyses and nutrient budgets allow evaluating the fertilization program on the long term ^[4].

General Sampling Instructions

- When plant development differs within a field, the field should be divided into different management areas with similar characteristics and a sample from each area should be taken. To facilitate interpretation, fields are best divided into the same areas as is done for soil samples.
- Randomly select plants throughout the field or management area and sample the correct plant parts ^[4, 13]. For plant parts and number of plant parts to sample, see Table 1.
- Collect the specific plant parts and place them into a clean paper bag ^[13]. Do not use plastic bags to avoid decay of samples. Do not use metal containers, because they may contaminate the samples and affect micronutrient results ^[4, 14].
- Do not collect samples during the hottest part of the day, particularly in summer ^[14].
- Do not take samples from dead, diseased, insect damaged, or mechanically injured plants ^[6, 13]. Also avoid plants from unusual areas in the field, including border areas and places where plants are under water stress or where nutrient availability is atypical ^[6].
- Dust or soil covered plant parts should also be avoided, especially when the samples are used for micronutrient analysis ^[6].
- Small amounts of dust can be removed by gently brushing the samples with a soft brush ^[13]. Alternatively, the samples may be cleaned with a damp cloth, but should not be rinsed or washed to prevent leaching of nutrients from the sample ^[13, 14].
- Deliver the samples immediately to the lab or use a one-day delivery service ^[13, 14]. If immediate delivery is not possible, air-dry the samples in the shade by placing the open bag in a clean, dust-free area ^[3, 4, 14]. Mix the samples frequently to avoid decay.
- Clearly label the bag, and provide the information required by the test lab ^[4].
- Follow the laboratory instructions for packaging and shipping.

- To determine the cause of visual symptoms or a suspected deficiency in one part of the field, two samples may be taken; one from the plants of interest, the other from adjoining normal plants in the same field or management area ^[6].

Table 1: Sampling procedure for major field crops

| Crop | Growth stage | Plant part to sample | Number of plants to sample |
|---------------------|--|---|-------------------------------------|
| Field Crops | | | |
| Alfalfa | 10% bloom | stems in the middle third of the plant | 40-60 stems from at least 30 plants |
| Dry Beans | Early growth | Petiole of fourth leaf from the growing tip | 40 |
| | Pre-bloom | Petiole of fourth leaf from the growing tip | 40 |
| | Late bloom | Petiole of fourth leaf from the growing tip | 40 |
| Corn | Early season (6-16 inches) | Whole plant | 20-30 |
| | Midgrowth (3-6 feet) | First fully developed leaf; third leaf from top | 15-25 |
| | Tasseling | Leaf opposite and below primary ear | 15-25 |
| | Silking | Leaf opposite and below primary ear | 15-25 |
| Cotton | Early squaring to late season | Third to fifth petiole from the terminal on the main stem | 30-40 |
| Rice | Early stages | Most recently fully expanded leaf (Y-leaf) | 50 |
| | Later stages | Most recently fully expanded leaf (Y-leaf) | 30-60 |
| Safflower | Prebloom | Stem, middle section | 40 |
| | First bud visible | Recently matured mid-stem leaves | 40 |
| Sunflower | Just before floret emergence | Most recently matured leaf (3rd or 4th from the terminal bud) | 20-30 |
| Processing Tomatoes | First bloom to 10% of fruits showing red color | Fourth leaf from the growing tip | 40 |
| Wheat and barley | 3-4 leaf | Whole plant | 50-100 |
| | Tillering | Top 3-4 leaves | 50-100 |
| | Jointing | Top 3-4 leaves | 50-100 |
| | Booting | Top 3-4 leaves | 50-100 |
| | Early heading (hard red wheat N only) | Flag leaf | 50-100 |

Sources: Alfalfa ^[9,12], dry beans ^[2], corn ^[6], cotton ^[1,11], rice ^[10], safflower ^[2,7], sunflower ^[2,4,7], tomatoes ^[7,8], wheat and barley ^[6]

Table 2: Sampling procedure for major vegetable and berry crops

| Crop | Growth stage | Plant part to sample | Number of plants to sample |
|-------------------|------------------------------|---|----------------------------|
| Vegetables | | | |
| Broccoli | First buds to heading | Recently matured leaf, typically 3-4 nodes down from the growing point | 20-60 |
| Carrot | Midgrowth (>4 inches high) | Most recently matured leaf or petiole | 20-30 |
| Cauliflower | Head initiation | Recently matured leaf, typically 3-4 nodes down from the growing point | 20 |
| | Preharvest | Recently matured leaf, typically 3-4 nodes down from the growing point | 20 |
| Celery | Mid-growth | Most recently matured leaf or petiole | 20 |
| | Preharvest | Most recently matured leaf or petiole | 20 |
| Melon | Early flower | Most recently matured leaf or petiole, typically 6th from the growing tip | 20-30 |
| | Early fruit set/bulking | Most recently matured leaf or petiole, typically 6th from the growing tip | 20-30 |
| | First harvest | Most recently matured leaf or petiole, typically 6th from the growing tip | 20-30 |
| Onion | Early season | Tallest leaf | 20-30 |
| | Midseason | Tallest leaf | 20-30 |
| | Late season | Tallest leaf | 20-30 |
| Potato | Early season | Petiole of fourth leaf from the growing tip | 40 |
| | Midseason | Petiole of fourth leaf from the growing tip | 40 |
| | Late season | Petiole of fourth leaf from the growing tip | 40 |
| Lettuce | Early heading to pre-harvest | Youngest wrapper leaf | 20-60 |
| Berries | | | |
| Strawberry | Preharvest | Young mature leaves | 30-40 |
| | Main harvest | Young mature leaves | 30-40 |

Sources: Broccoli, cauliflower, celery and lettuce ^[5, 6], carrot ^[2,6], melon ^[2,7], onion ^[8], potato ^[7,16], strawberry ^[15]

References

1. Bassett, D.M., MacKenzie, A.J., 1976. Plant analysis as a guide to cotton fertilization. In: Reisenauer, H.M. (Ed.). Soil and Plant-Tissue Testing in California. University of California Cooperative Extension Bulletin 1879. pp. 16-17.
2. California Plant Health Association, 2002. Western Fertilizer Handbook 9th edition. Interstate Publishers, Inc.
3. Fernandez, F.G., Hoefft, R.G., 2012. Managing soil pH and crop nutrients. University of Illinois Extension - Illinois Agronomy Handbook. Available online at: <http://extension.cropsci.illinois.edu/handbook/pdfs/chapter08.pdf>
4. Flynn, R., Ball, S.T., Baker, R.D., 1999. Sampling for plant tissue analysis. New Mexico State University Cooperative Extension Service. Available online at: http://aces.nmsu.edu/pubs/_a/A123.pdf
5. Hartz, T.K., 2007. Efficient nitrogen management for cool-season vegetables. Available online at: http://vric.ucdavis.edu/pdf/fertilization_EfficientNitrogenManagementforCoolSeasonVegetable2007.pdf
6. Jones Jr., J.B., 1998. Field sampling procedures for conducting a plant analysis. In: Kalra, Y.P. (Ed.). Handbook of Reference Methods for Plant Analysis. CRC Press, Boca Raton. pp. 25-35.
7. Lorenz, O.A., Tyler, K.B., 1976. Plant tissue analysis of vegetable crops. In: Reisenauer, H.M. (Ed.). Soil and Plant-Tissue Testing in California. University of California Cooperative Extension Bulletin 1879. pp. 24-29.
8. Maynard, D.N., Hochmuth, G.J., 2007. Knott's Handbook for Vegetable Growers. John Wiley & Sons, Inc., Hoboken, NJ.
9. Meyer, R.D., Marcum, D.B., Schmierer, J.L., 1998. Do I need fertilizer? - Ask the plant! Proceedings of the 28th California/Nevada Alfalfa Symposium. Available online at: <http://alfalfa.ucdavis.edu/+symposium/proceedings/1998/98-159.pdf>
10. Mikkelsen, D.S., 1976. Diagnostic plant analysis for rice. In: Reisenauer, H.M. (Ed.). Soil and Plant-Tissue Testing in California. University of California Cooperative Extension Bulletin 1879. pp. 30-31.
11. Reddy, K.R., Hodges, H.F., Varco, J., 2000. Potassium nutrition of cotton. Mississippi Agricultural and Forestry Experiment Station Bulletin 1049. Available online at <http://msucares.com/pubs/bulletins/b1094.pdf>
12. Sallee, W.R., Ulrich, A., Martin, W.E., Krantz, B.A., 1959. High phosphorus for alfalfa: Plant analysis used to evaluate phosphorus status of alfalfa fields as guide to fertilizing for better yields and returns. California Agriculture 13(8), 7-8.
13. Schwab, G.J., Lee, C.D., Pearce, R., 2007. Sampling plant tissue for nutrient analysis. University of Kentucky Cooperative Extension Service. Available online at: <http://www.ca.uky.edu/agc/pubs/agr/agr92/agr92.pdf>
14. Thom, W.O., Brown, J.R., Plank, C.O., 1991. Sampling for corn plant tissue analysis. Iowa State University Extension. Available online at: <http://www.extension.iastate.edu/Publications/NCH15.pdf>
15. Ulrich, A., 1976. Plant tissue analysis as a guide in fertilizing crops. In: Reisenauer, H.M. (Ed.). Soil and Plant-Tissue Testing in California. University of California Cooperative Extension Bulletin 1879. pp. 6-8
16. Voss, R.E., 2004. Reducing fertilizer needs of potato with new varieties and clonal strains of existing varieties. FREP final report. Available online at: <http://www.cdfa.ca.gov/is/ffldrs/frep/pdfs/completedprojects/00-0514Voss2004.pdf>

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